

Selective oncological theragnostic based on radioactively labeled exosomes

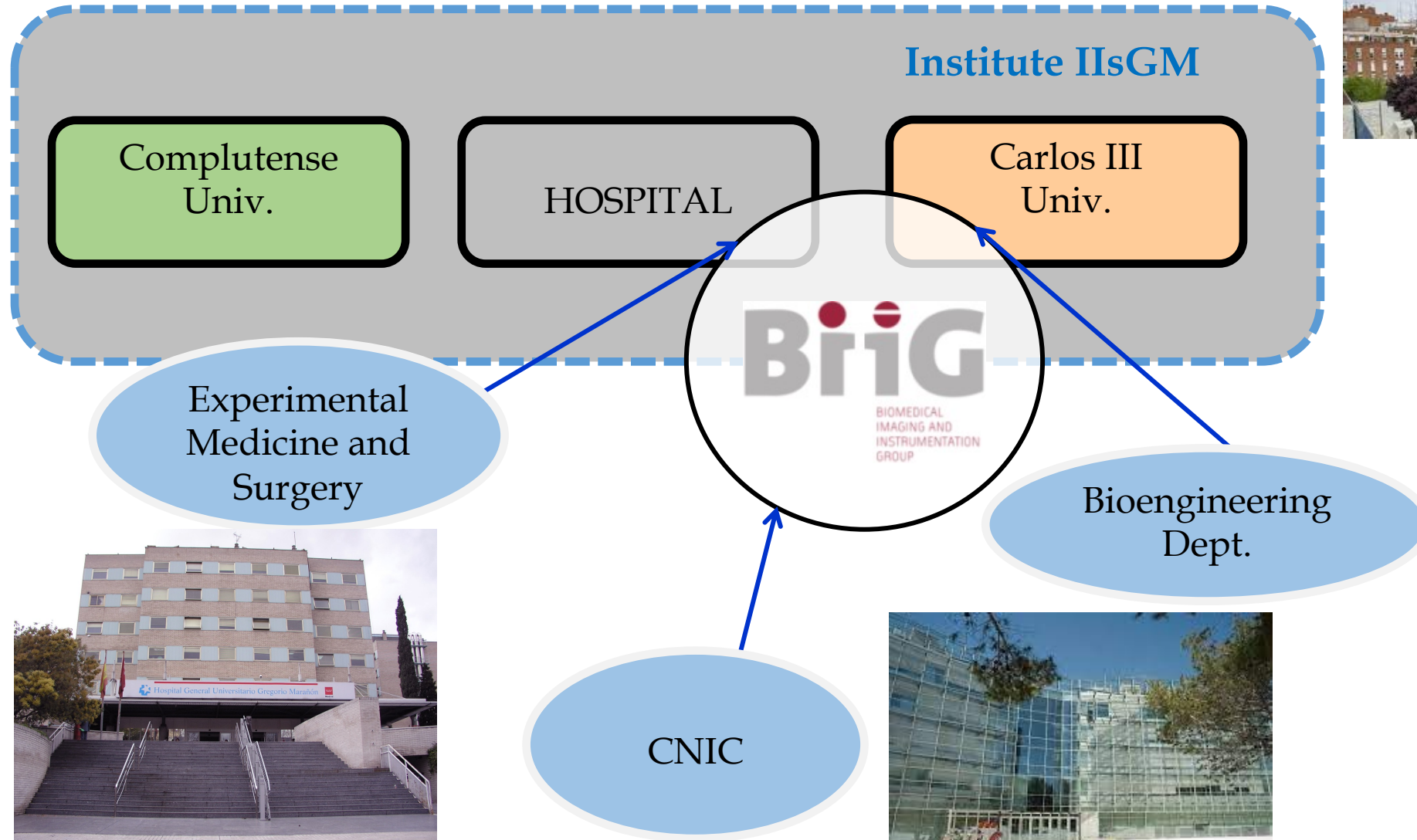
Beatriz Salinas Rodríguez

“Molecular probes” Lab Head
Biomedical imaging and instrumentation Group
Instituto de investigaciones Sanitarias Gregorio Marañón.

Biomedical engineering dept, Universidad Carlos III Madrid.

Advanced Imaging Unit, National Center of Cardiovascular Diseases

Biomedical Imaging & Instrumentation Group

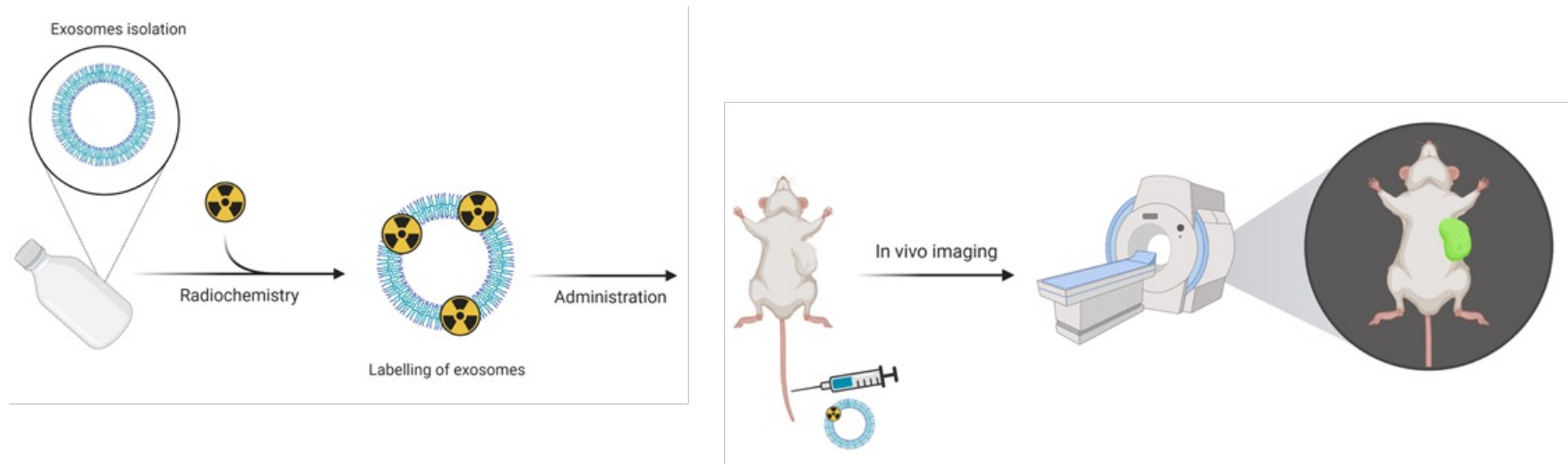


Molecular Probes Lab



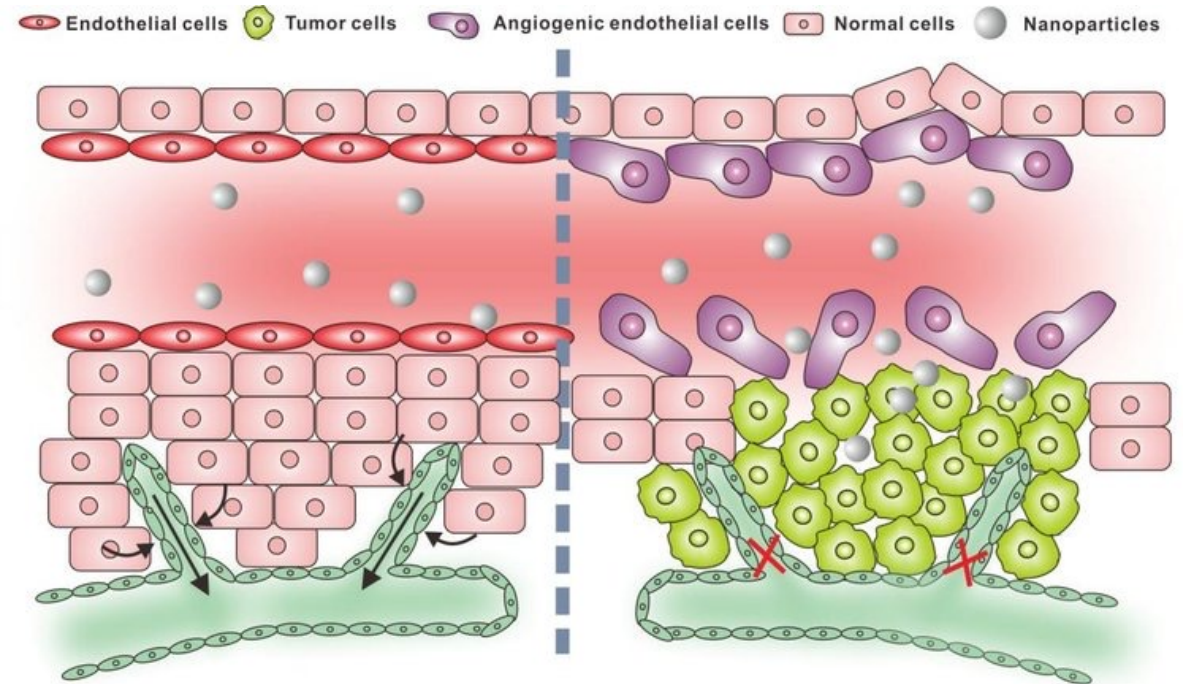
PRISMAP project

Development of new radiotheragnostic agents based on natural nanoparticles (exosomes) radioactively labeled with the novel therapeutic and diagnostic isotopes Terbium 161.



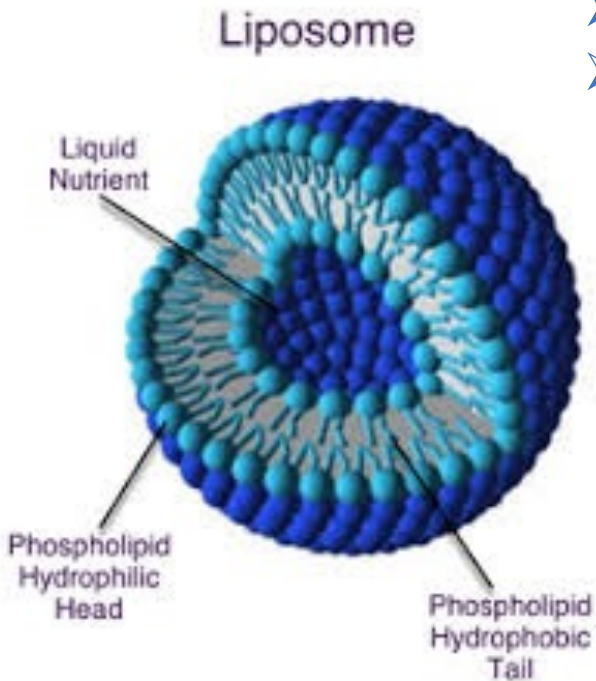
Nanoparticles theragnostics

- Large surface-to-volume ratios
- An arsenal of surface functionalization possibilities (functional groups)
- Capacity to carry large amounts of cargo
- Enhance permeability retention effect (EPR)



Liposomes and nanotechnology

STRUCTURE AND APPLICATIONS



- Drug cargo
- Superficial modification

FROM BENCH TO BEDSIDE



Pubmed: liposomes + preclinic +

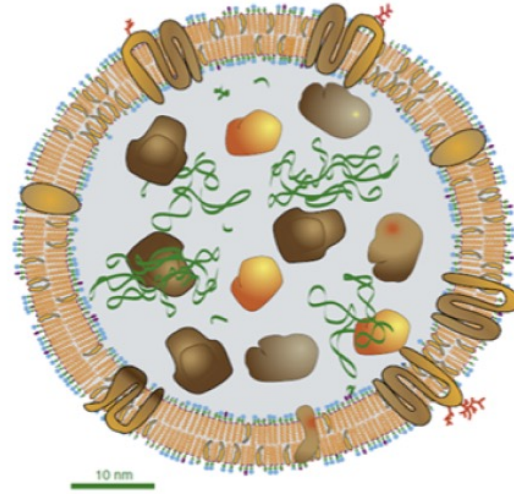
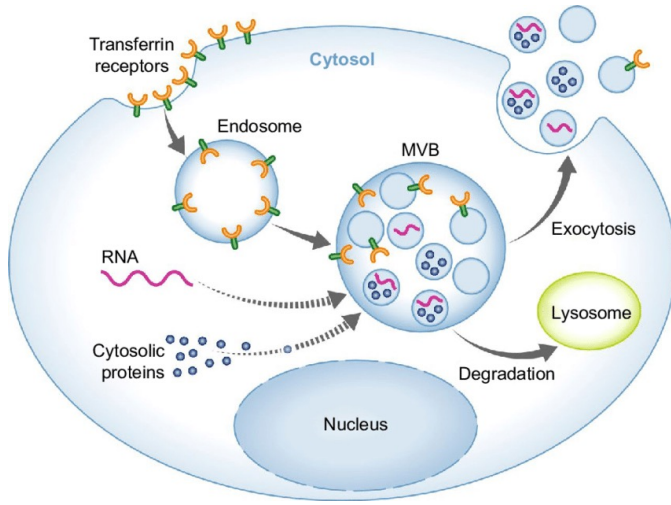
- Diagnostic : 12.873
- Therapy: 22.131

Table 1. Clinically used liposome-based products.

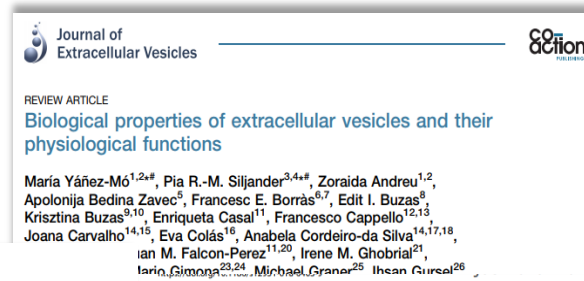
SN	Clinical Products (Approval Year)	Administration	Active Agent	Lipid/Lipid:Drug Molar Ratio	Indication	Company
1.	Doxil® (1995)	i.v.	Doxorubicin	HSPC:Cholesterol:PEG 2000-DSPE (56:39:5 molar ratio)	Ovarian, breast cancer, Kaposi's sarcoma	Sequus Pharmaceuticals
2.	DaunoXome® (1996)	i.v.	Daunorubicin	DSPC and Cholesterol (2:1 molar ratio)	AIDS-related Kaposi's sarcoma	NoStar Pharmaceuticals
3.	Depocyt® (1999)	Spinal	Cytarabine/Ara-C	DOPC, DPPG, Cholesterol and Triolein	Neoplastic meningitis	SkyPharma Inc.
4.	Myocet® (2000)	i.v.	Doxorubicin	EPC:Cholesterol (55:45 molar ratio)	Combination therapy with cyclophosphamide in metastatic breast cancer	Elan Pharmaceuticals
5.	Mepact® (2004)	i.v.	Mifamurtide	DOPS:FOPC (3:7 molar ratio)	High-grade, resectable, non-metastatic osteosarcoma	Takeda Pharmaceutical Limited
6.	Marqibo® (2012)	i.v.	Vincristine	SM:Cholesterol (60:40 molar ratio)	Acute lymphoblastic leukaemia	Talon Therapeutics, Inc.
7.	Onivyde™ (2015)	i.v.	Irinotecan	DSPC:MPEG-2000:DSPE (3:2:0.015 molar ratio)	Combination therapy with fluorouracil and leucovorin in metastatic adenocarcinoma of the pancreas	Merrimack Pharmaceuticals Inc.
8.	Abelcet® (1995)	i.v.	Amphotericin B	DMPC:DMPG (7:3 molar ratio)	Invasive severe fungal infections	Sigma-Tau Pharmaceuticals
9.	Ambisome® (1997)	i.v.	Amphotericin B	HSPC:DSPG:Cholesterol:Amphotericin B (2:0.8:1:0.4 molar ratio)	Presumed fungal infections	Astellas Pharma
10.	Amphotec® (1996)	i.v.	Amphotericin B	Cholesteryl sulphate:Amphotericin B (1:1 molar ratio)	Severe fungal infections	Ben Venue Laboratories Inc.
11.	Visudyne® (2000)	i.v.	Verteporfin	Verteporfin:DMPC and EPG (1:8 molar ratio)	Choroidal neovascularisation	Novartis
12.	DepoDur™ (2004)	Epidural	Morphine sulfate	DOPC, DPPG, Cholesterol and Triolein	Pain management	SkyPharma Inc.
13.	Esparel® (2011)	i.v.	Bupivacaine	DEPC, DPPG, Cholesterol and Tricaprylin	Pain management	Pacira Pharmaceuticals, Inc.
14.	Epaxal® (1993)	i.m.	Inactivated hepatitis A virus (strain HGSB)	DOPC:DOPE (75:25 molar ratio)	Hepatitis A	Crucell, Berna Biotech
15.	Inflexal® V (1997)	i.m.	Inactivated hemagglutinin of Influenza virus strains A and B	DOPC:DOPE (75:25 molar ratio)	Influenza	Crucell, Berna Biotech

i.v. (intravenous); i.m. (intramuscular); HSPC (hydrogenated soy phosphatidylcholine); PEG (polyethylene glycol); DSPE (distearoyl-sn-glycero-phosphoethanolamine); DSPC (distearoylphosphatidylcholine); DOPC (dioleoylphosphatidylcholine); DPPG (dipalmitoylphosphatidylglycerol); EPC (egg phosphatidylcholine); DOPS (dioleoylphosphatidylserine); POPC (palmitoyl-oleoylphosphatidylcholine); SM (sphingomyelin); MPEG (methoxy polyethylene glycol); DMPC (dimyristoyl phosphatidylcholine); DMPG (dimyristoyl phosphatidylglycerol); DSPG (distearoylphosphatidylglycerol); DEPC (dierucoylphosphatidylcholine); DOPE (dioleoyl-sn-glycero-phosphoethanolamine).

Exosomes (SEVs) : natural liposomes



- Extracellular vesicles endosomal origin
- nanometric size: 50 – 150 nm
- Extracellular communication.
- **Structure: lipid bilayer (liposomes)**



Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET

Héctor Peinado¹, Maša Alečković², Simon Lavotshkin³, Irina Matei¹, Bruno Costa-Silva^{1,4}, Gema Moreno-Bueno⁵, Marta Hergueta-Redondo⁵, Caitlin Williams⁵, Guillermo Garcia-Santos⁵, Cyrus M Ghajar⁶, Ayuko Nitadori-Hoshino⁷, Caitlin Hoffman⁸, Karen Badal¹, Benjamin A Garcia⁹, Margaret K Callahan⁹, Jianda Yuan⁹, Vilma R Martins⁴, Roberto Soto¹⁰, Rosendo N Kozlov¹¹, Mary S Reed¹², Todd D Wolchok^{5,9,13}, Paul R Chapman^{8,13}, Yibin Kang^{14,15}



Biological functions:

- Cell-cell communication
- Disease progression
- Tumor evolution

Natural NPs with biological function

Goat milk exosomes as natural liposomes

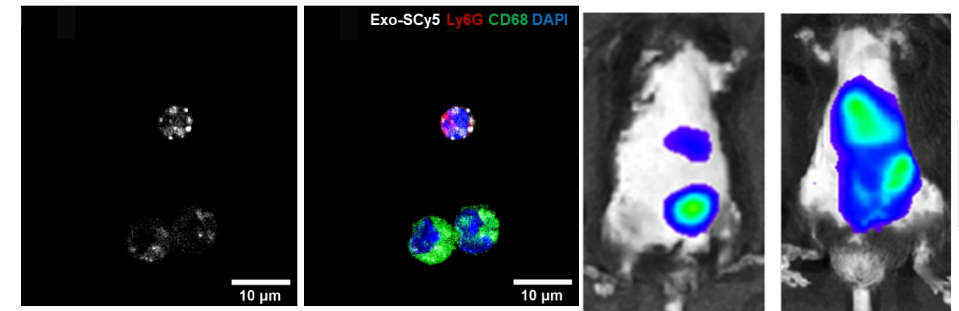
Advantages

- Vs synthetic nanoparticles (liposomes): Biocompatibility, biodegradability
- Vs other exosomes:
 - *Economic*
 - *Scalable*
 - *Robust in degrading conditions (Pieters, B.C., et al., 2015)*
 - *Non-immunogenicity and cross-species tolerance (Munagala, R. et al., 2016)*
 - *Resistant*



Applications:

- ❖ **Drug delivery:** structure like liposome
- ❖ **Early detection:** biomarkers

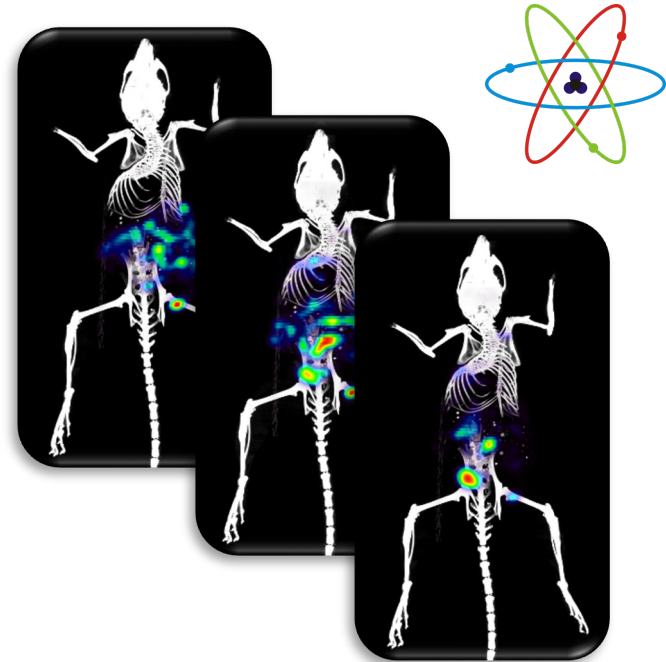


MOLECULAR IMAGING

Optical imaging

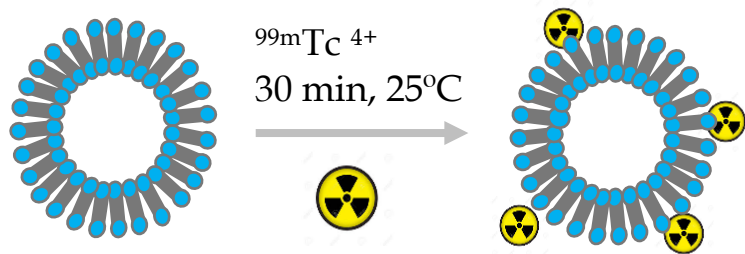


Nuclear imaging

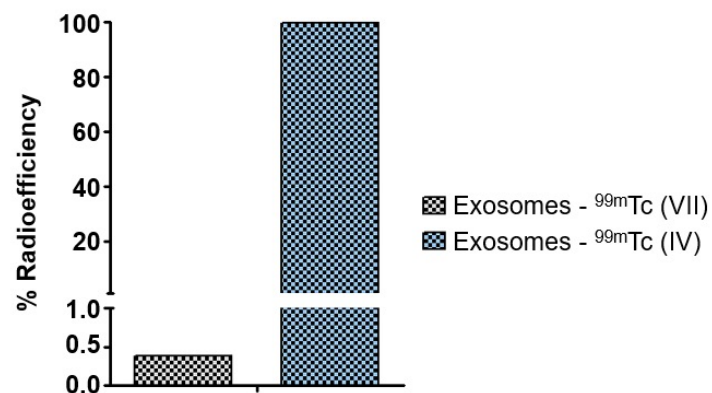




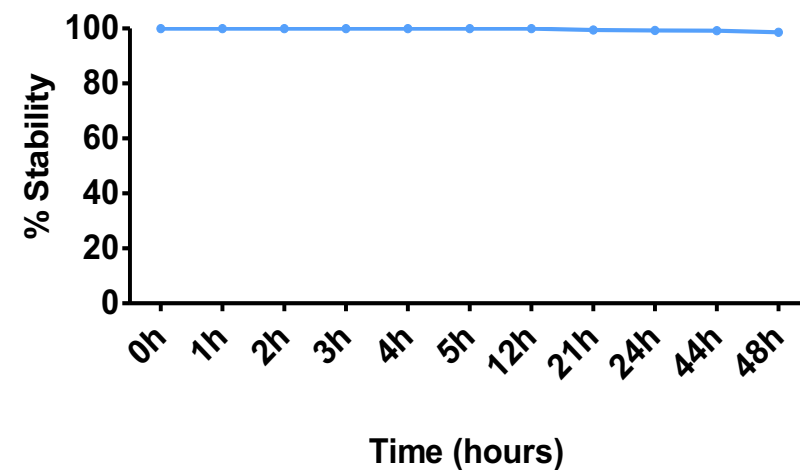
A.- Radiosynthesis



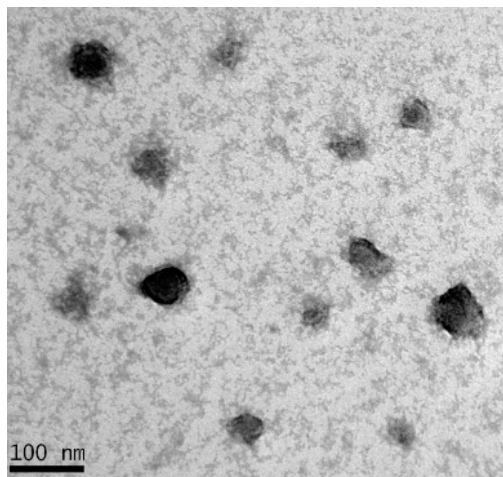
B.- Passive incorporation



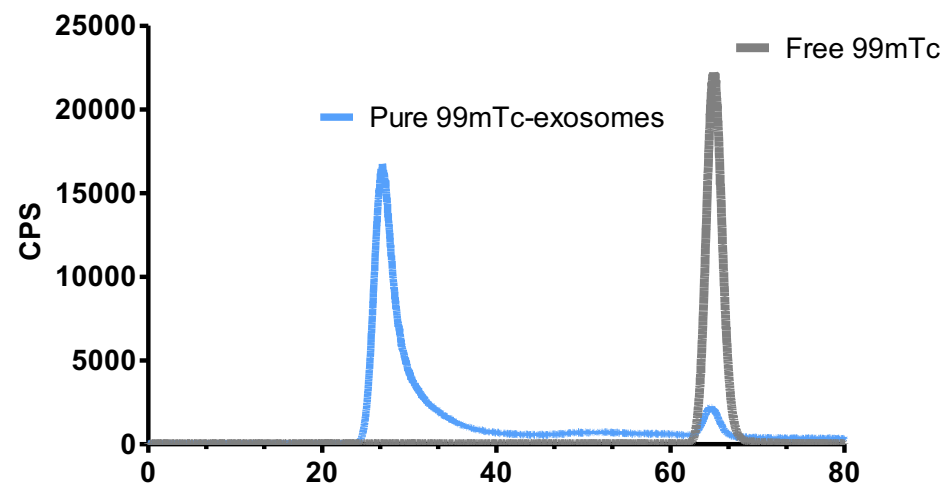
C.- Stability (in vitro)



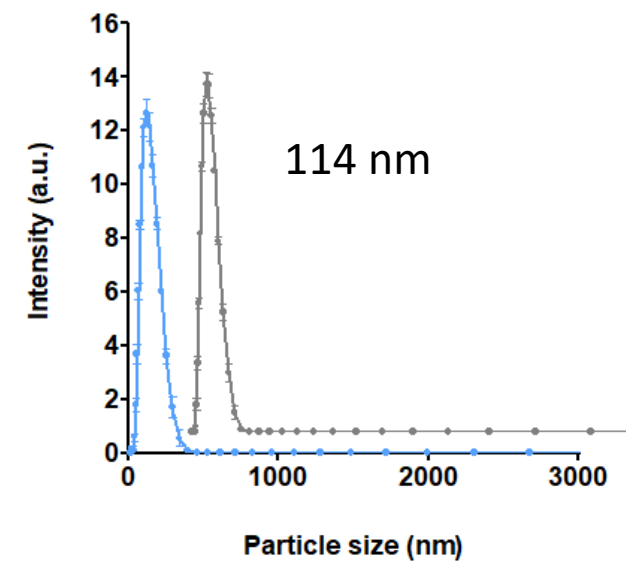
D.- TEM (microscopy)



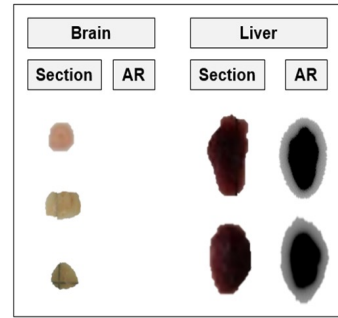
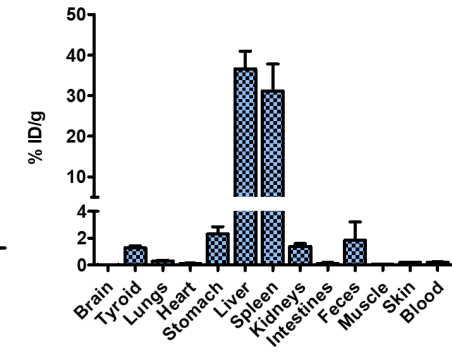
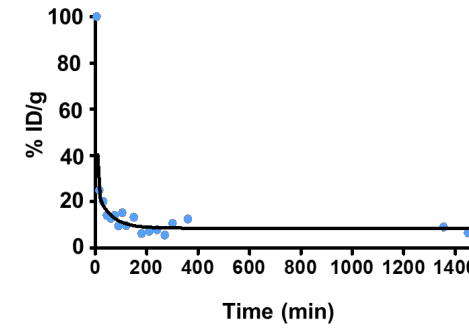
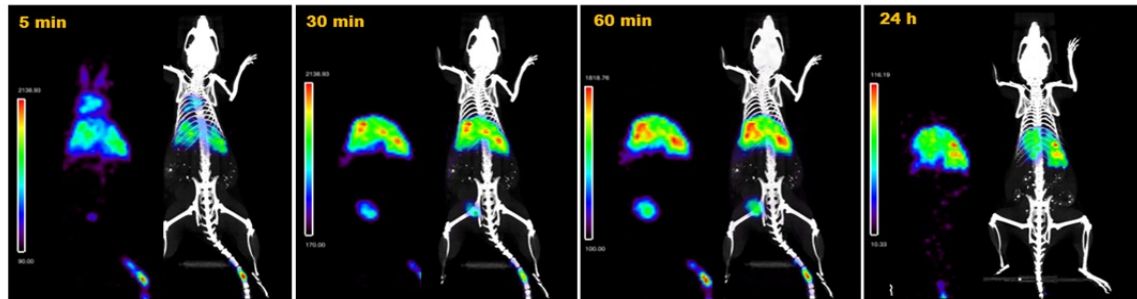
E.- RadioHPLC (purity)



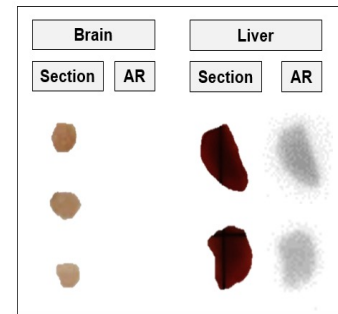
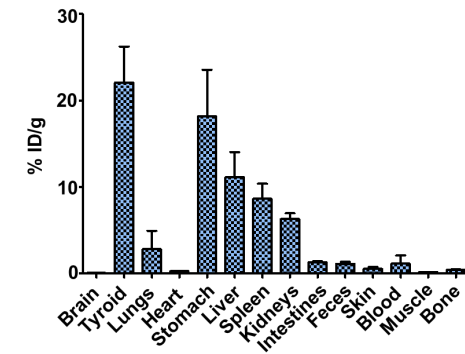
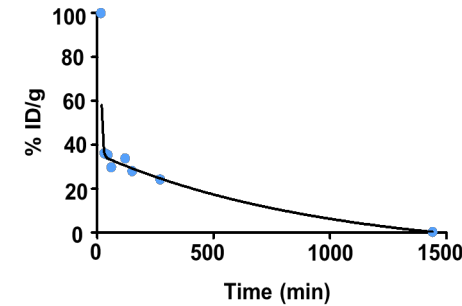
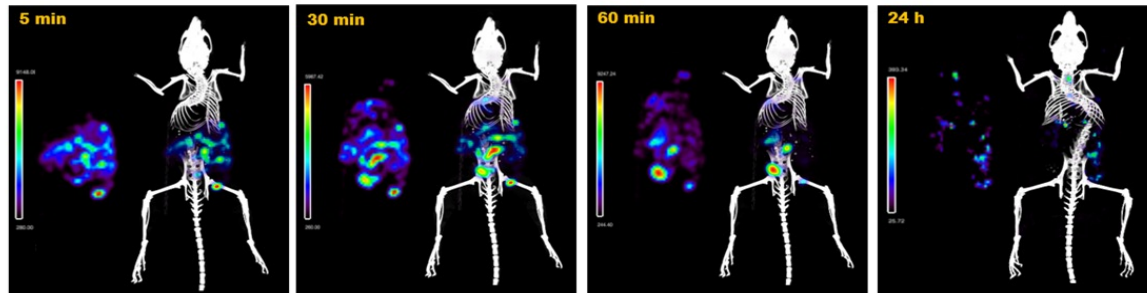
F.- DLS (size)



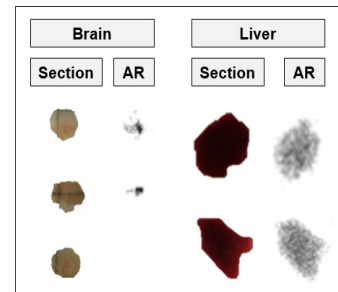
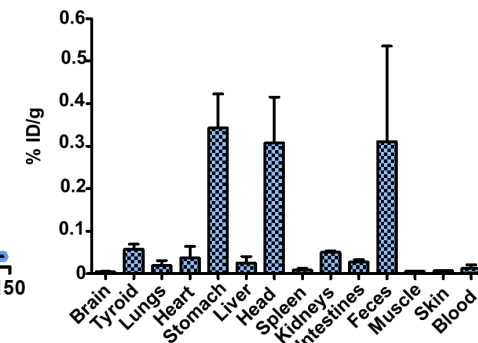
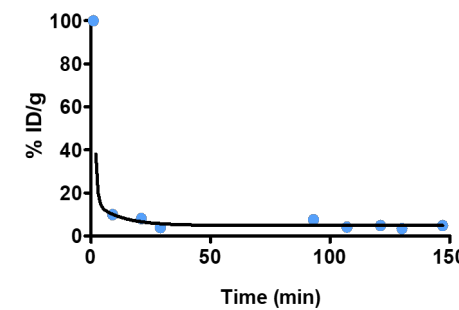
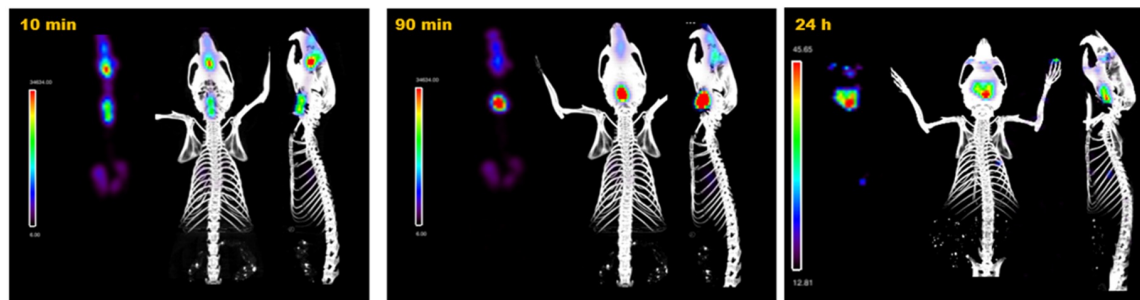
► Intravenous administration



► Intraperitoneal administration

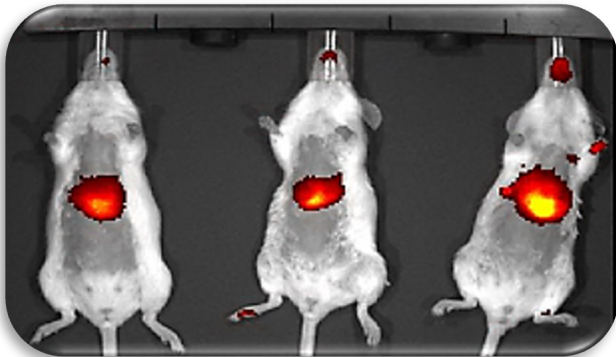


► Intranasal administration

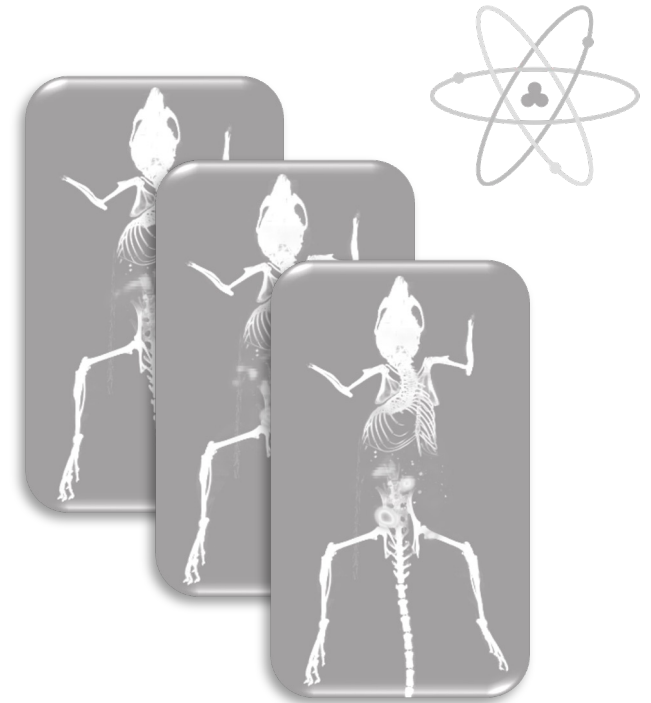


MOLECULAR IMAGING

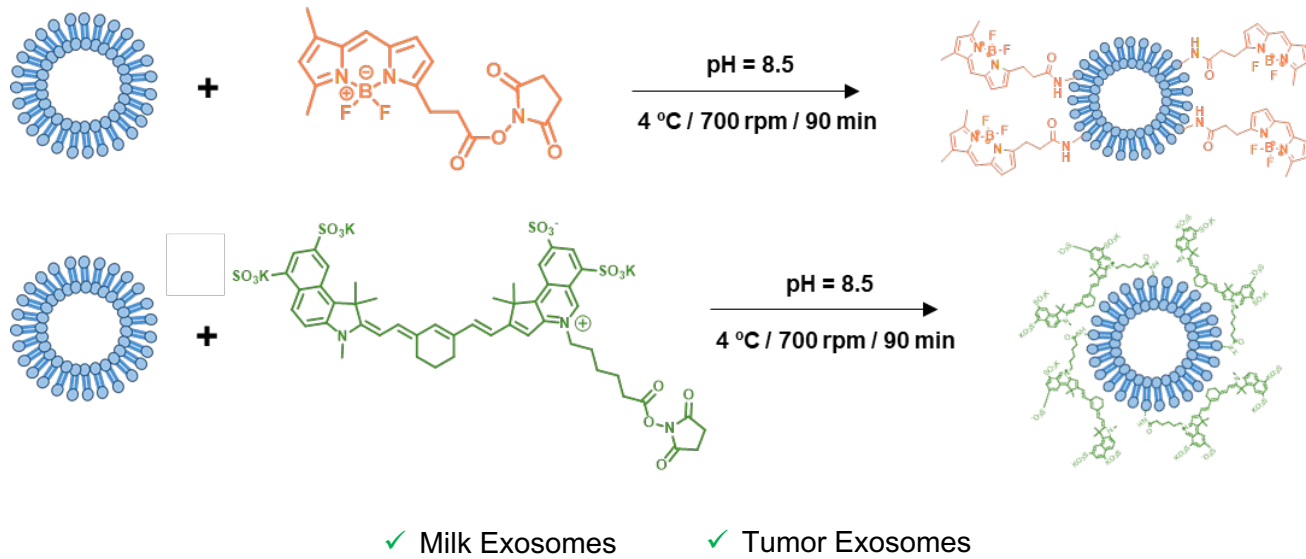
Optical imaging



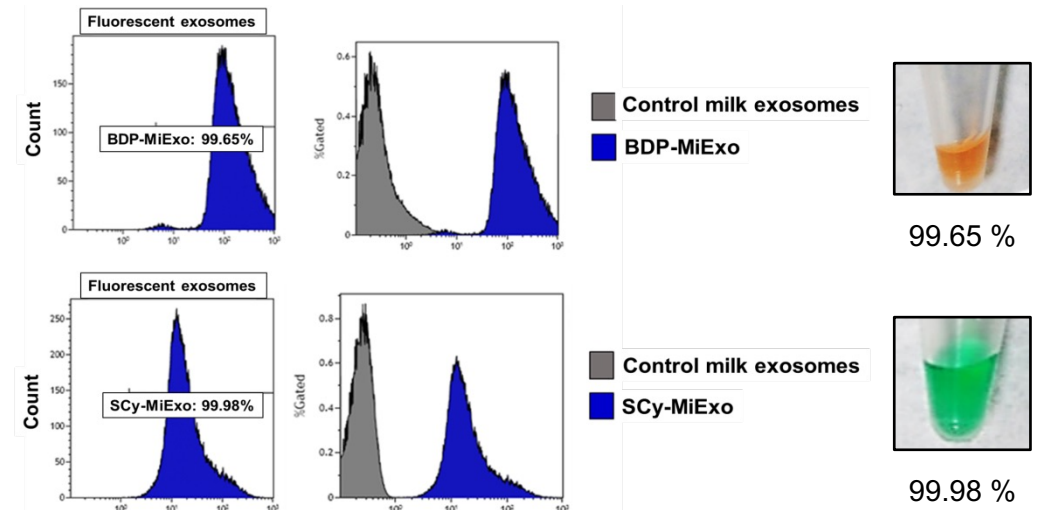
Nuclear imaging



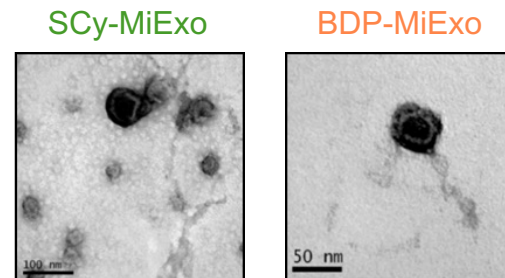
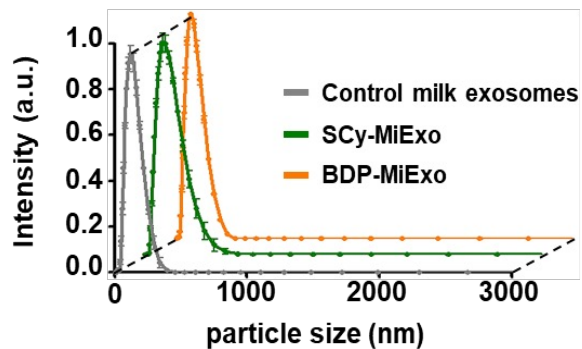
A.- Reaction



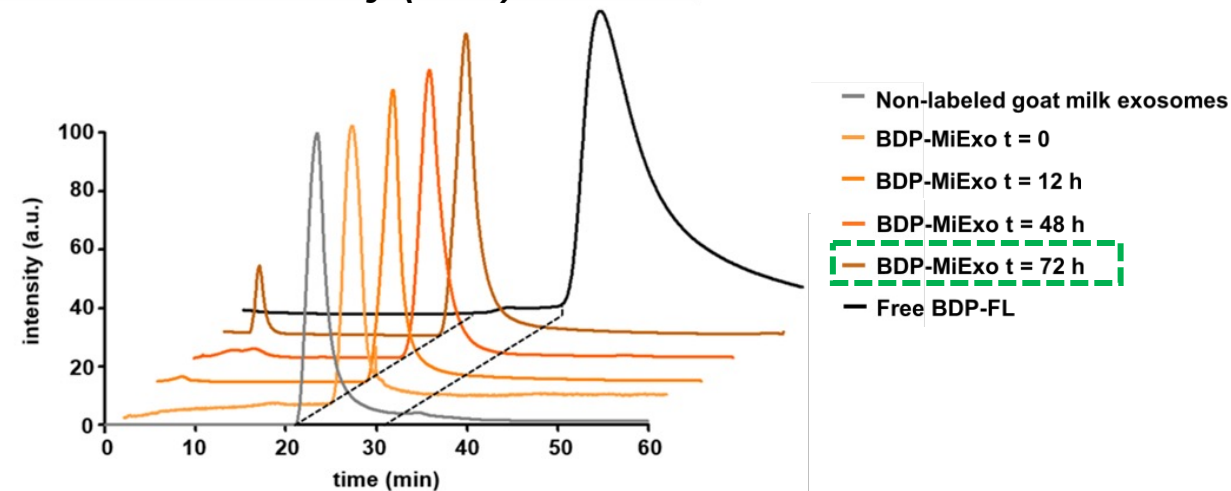
B.- Flow cytometry and yield of labelling



C.- Physicochemical characterization



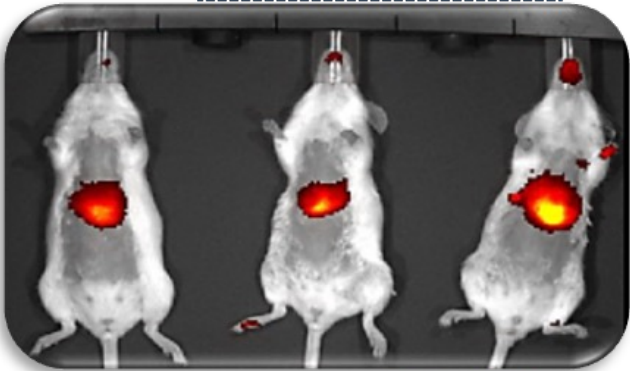
D.- In vitro stability (PBS):



MOLECULAR IMAGING

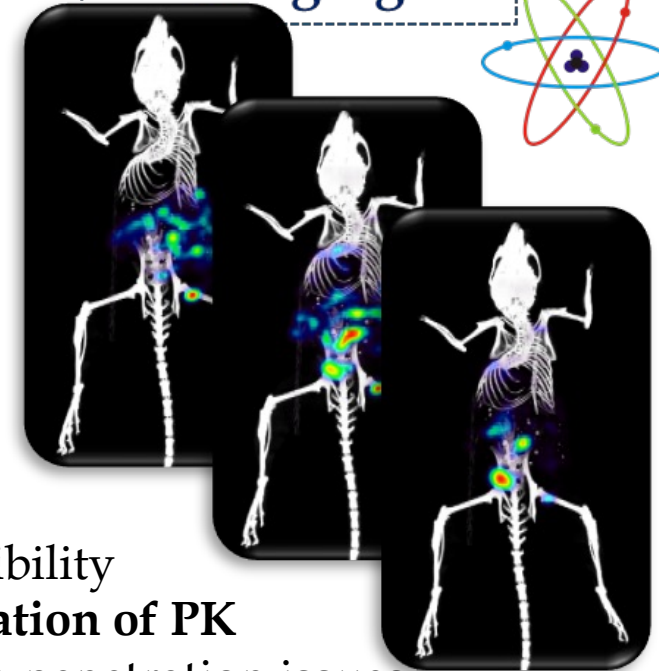
Dual probe

Optical
imaging



- High sensibility
- **Visualization (in vitro, histology)**
- Non-ionizing radiation

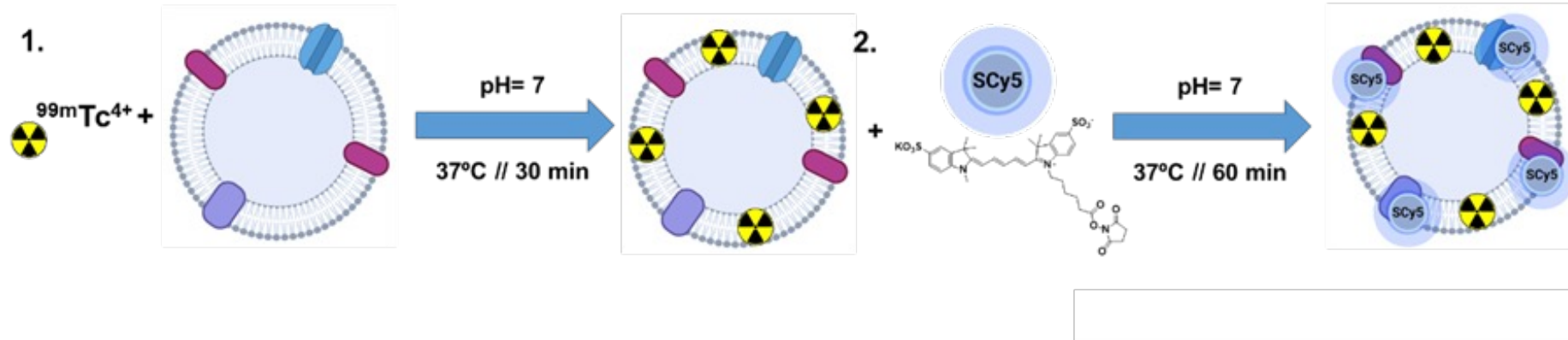
Nuclear
imaging



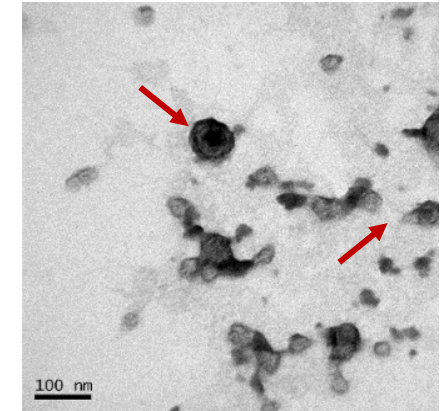
- High sensibility
- **Quantification of PK**
- Non depth penetration issues

Dual labelling and characterization

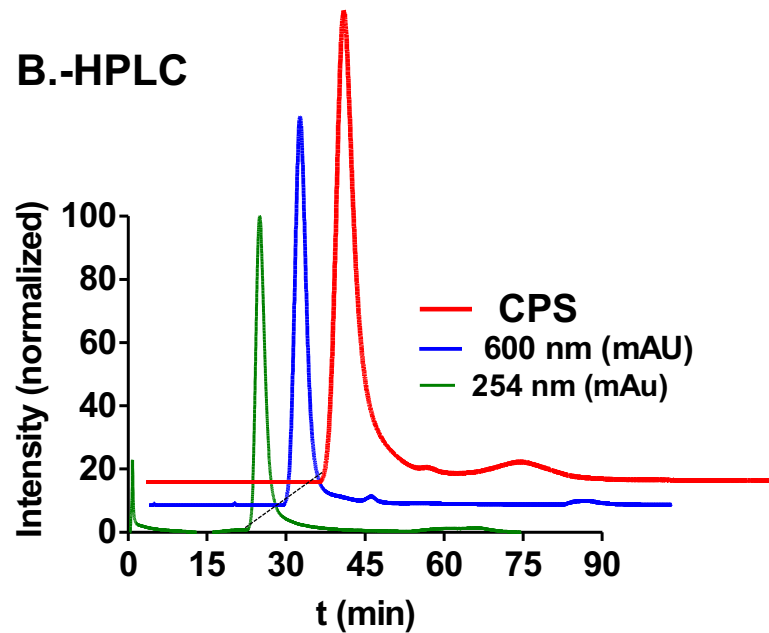
Reaction



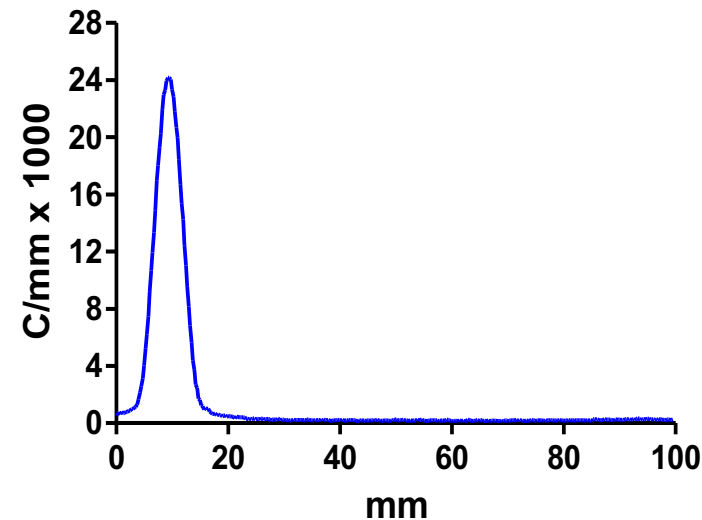
A.-TEM



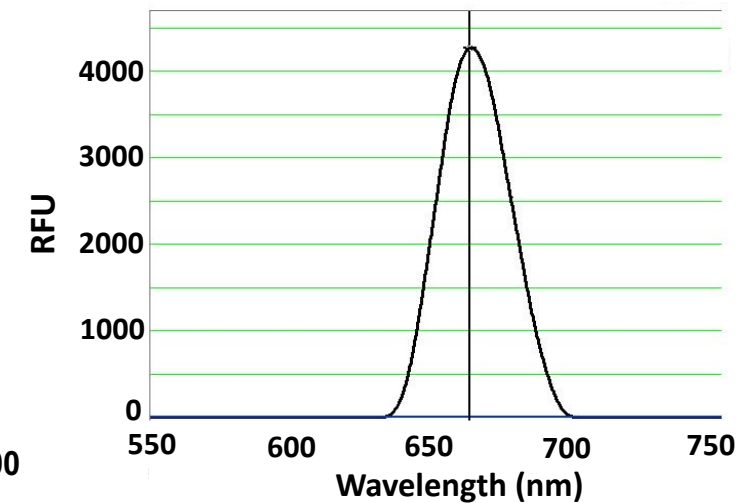
B.-HPLC



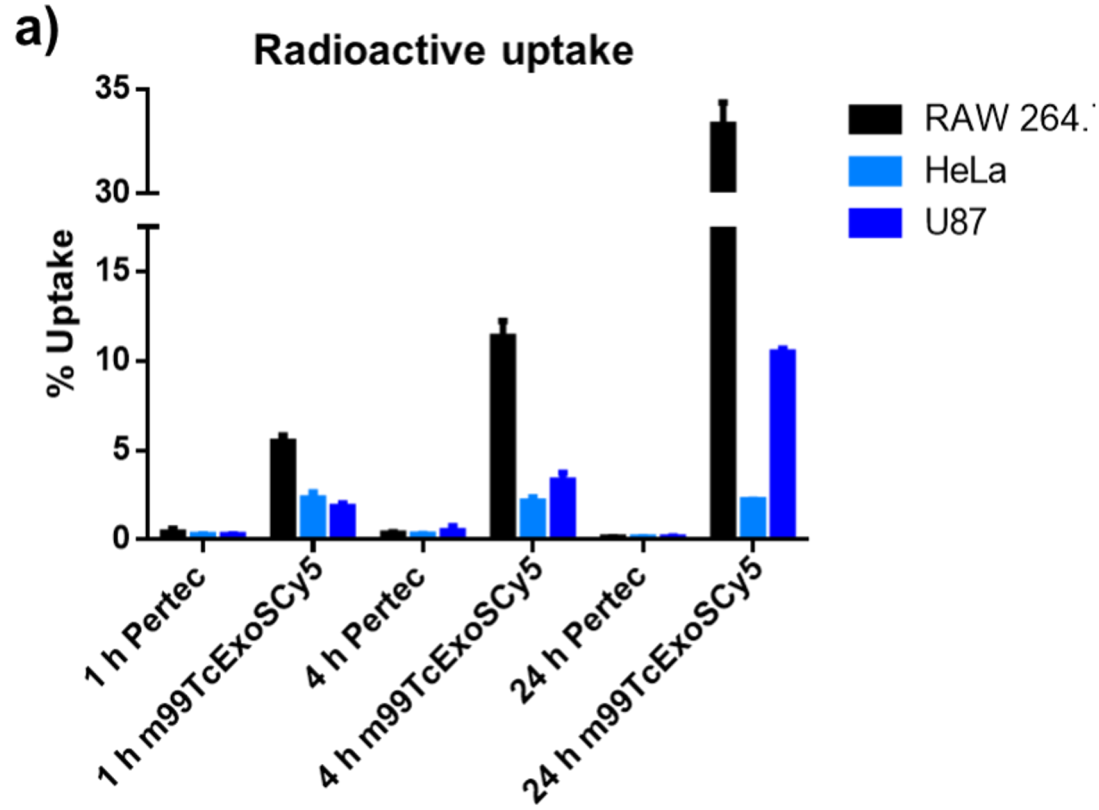
C.-iTLC



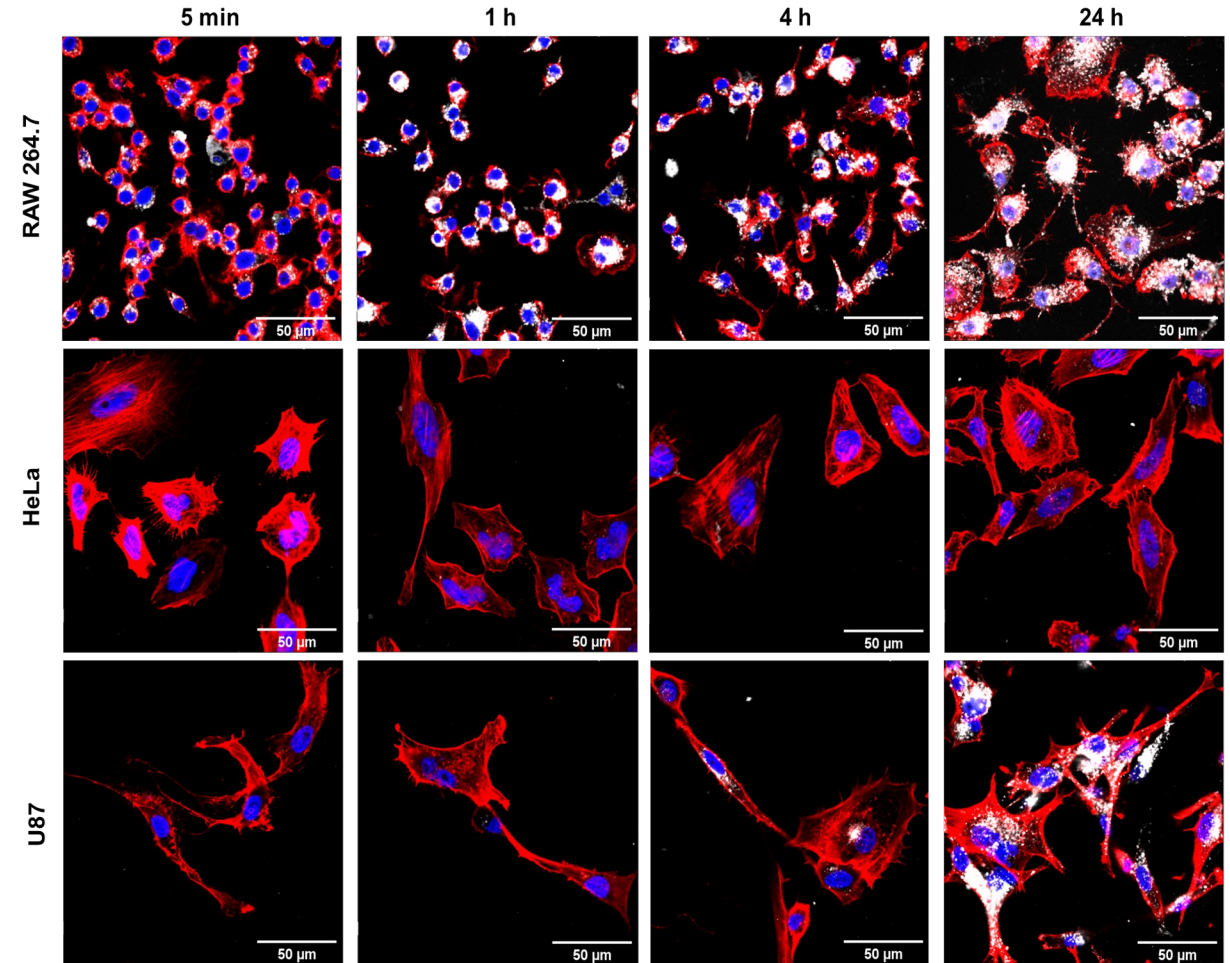
D.-Fluorescence

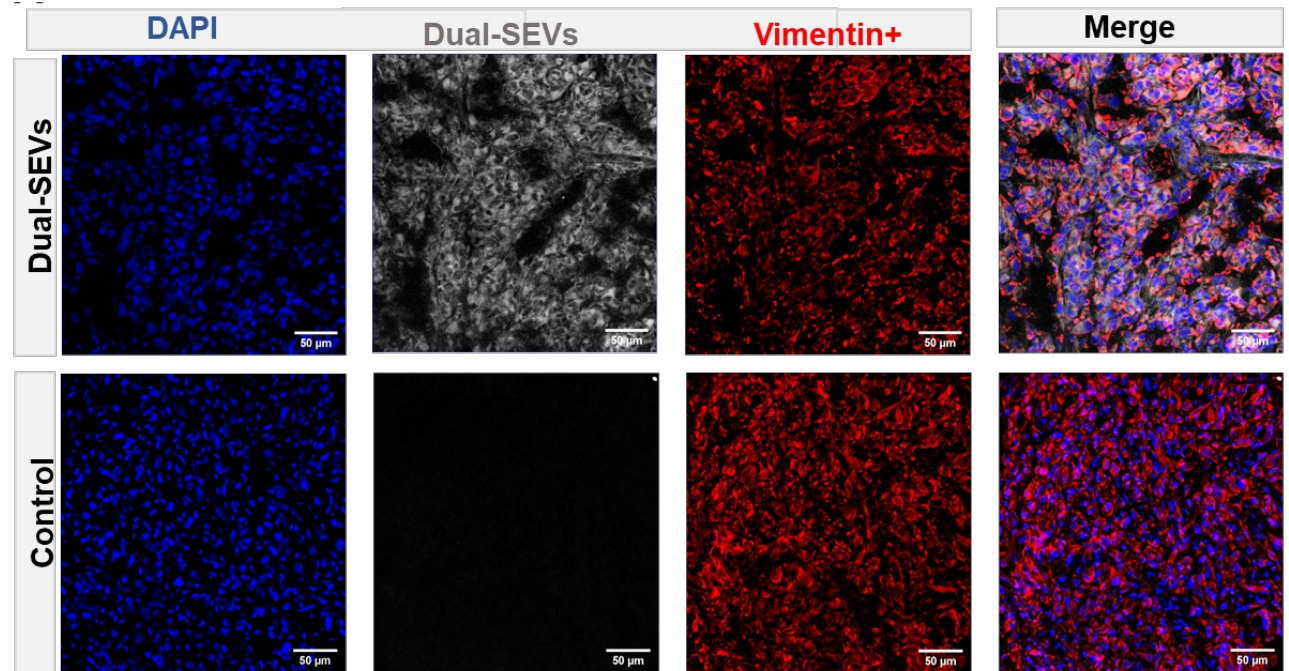
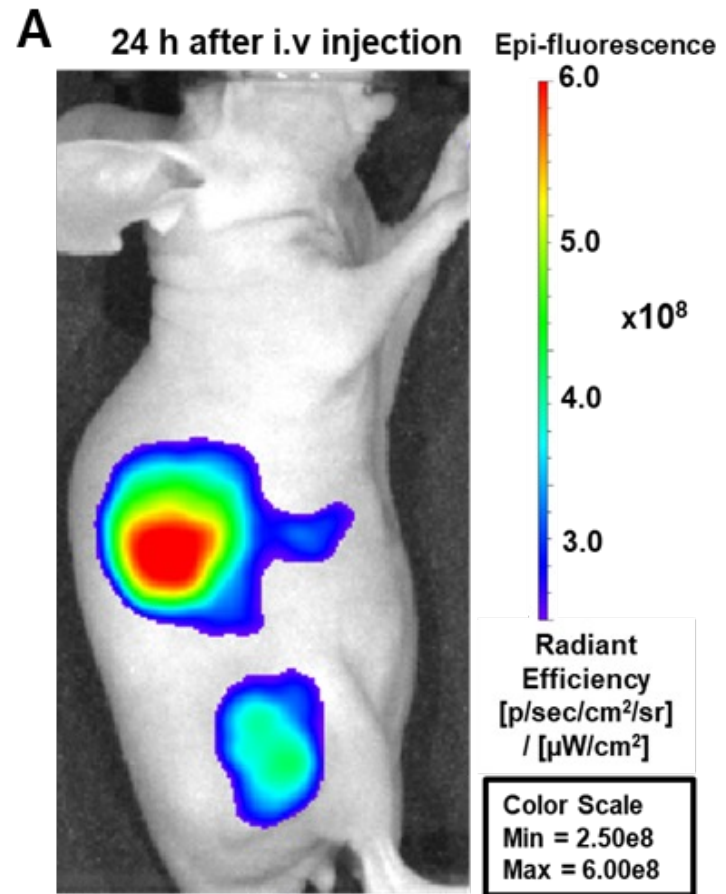


In vitro assessment (Radioactive and optical)

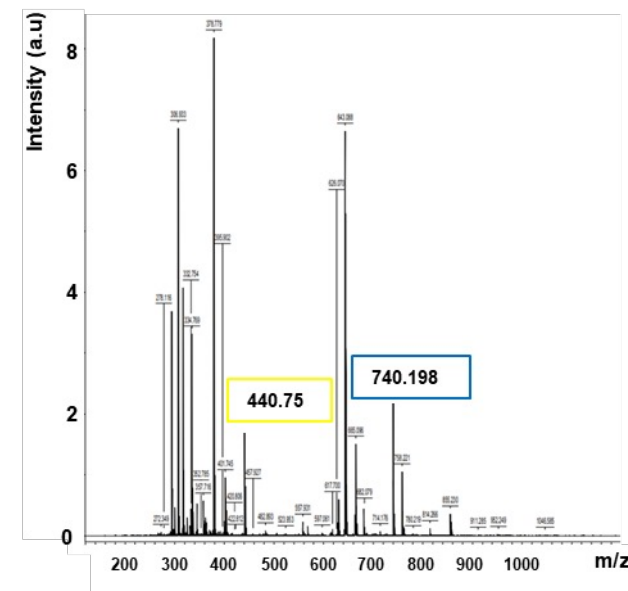
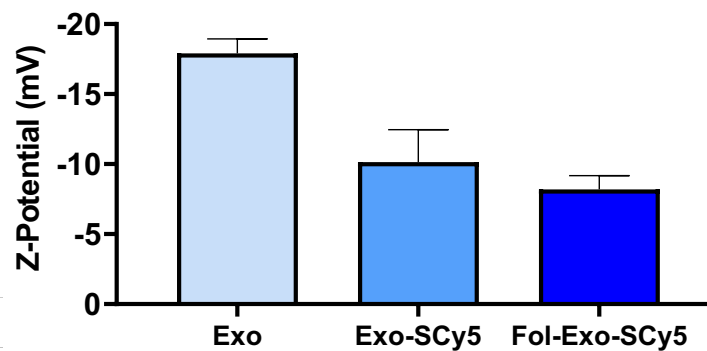
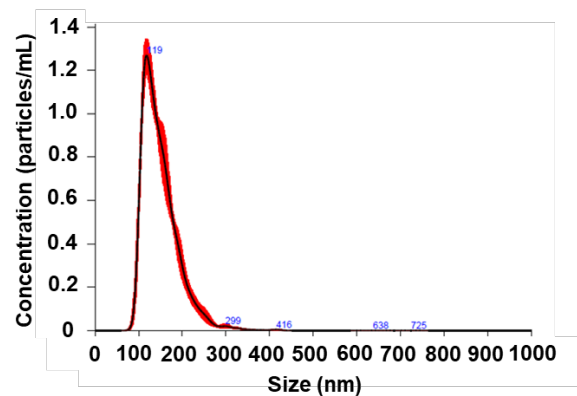
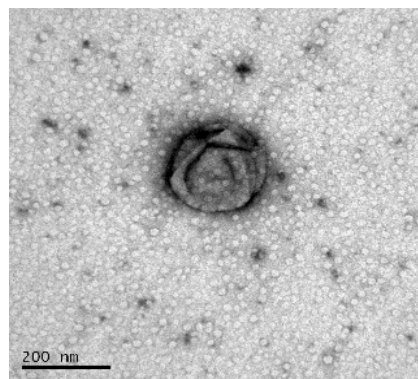
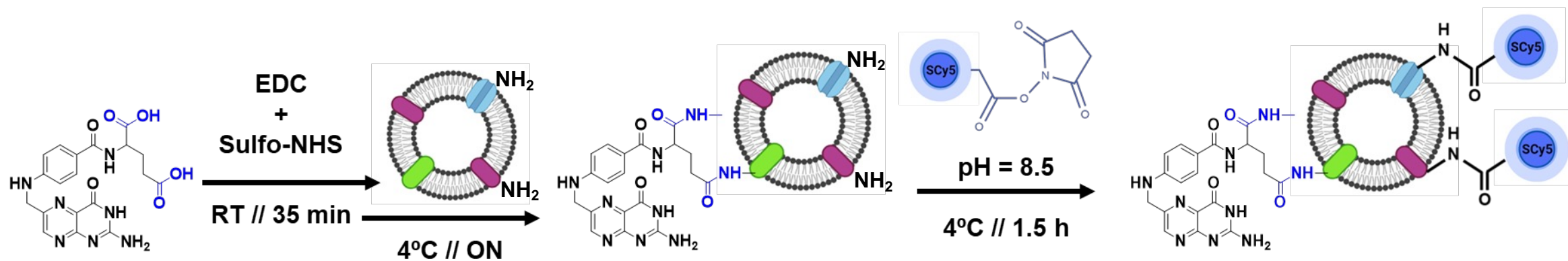


b) Confocal

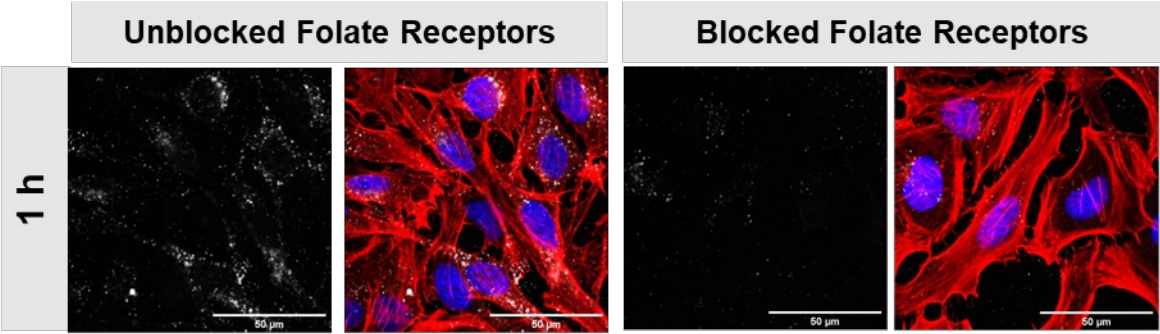
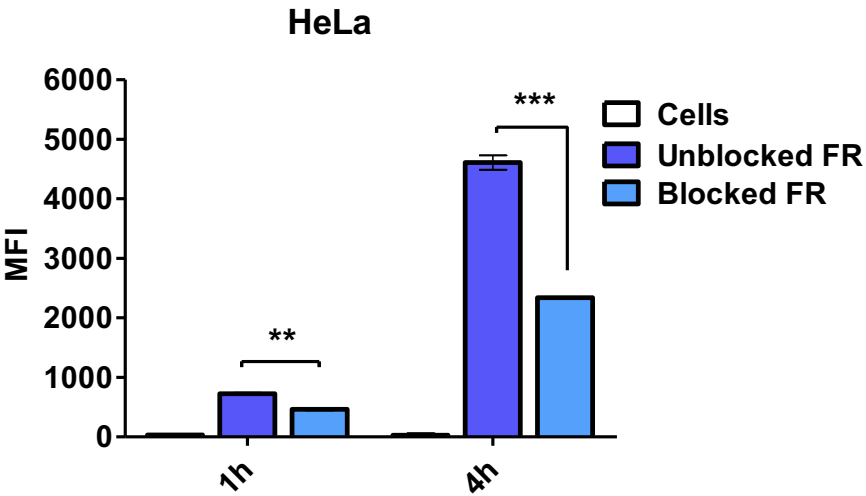




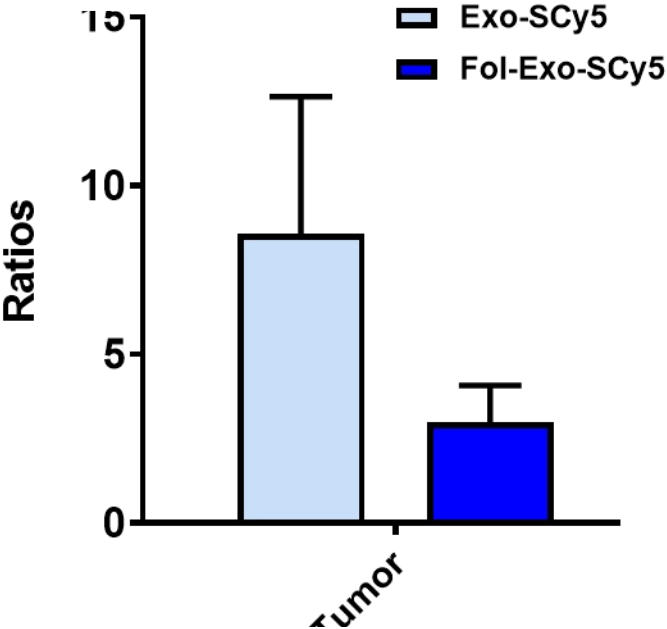
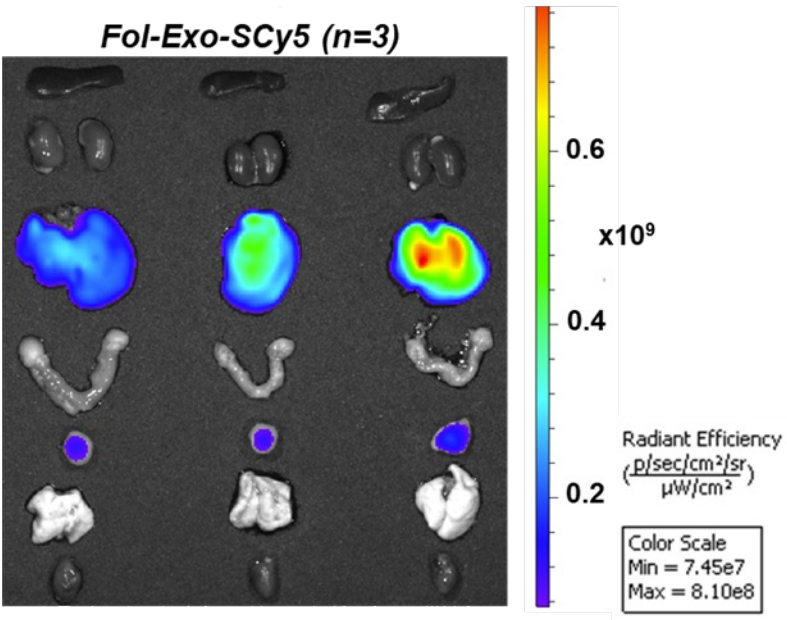
Exosome engineering_ Folic acid



A.- In vitro evaluation



B.- In vivo validation

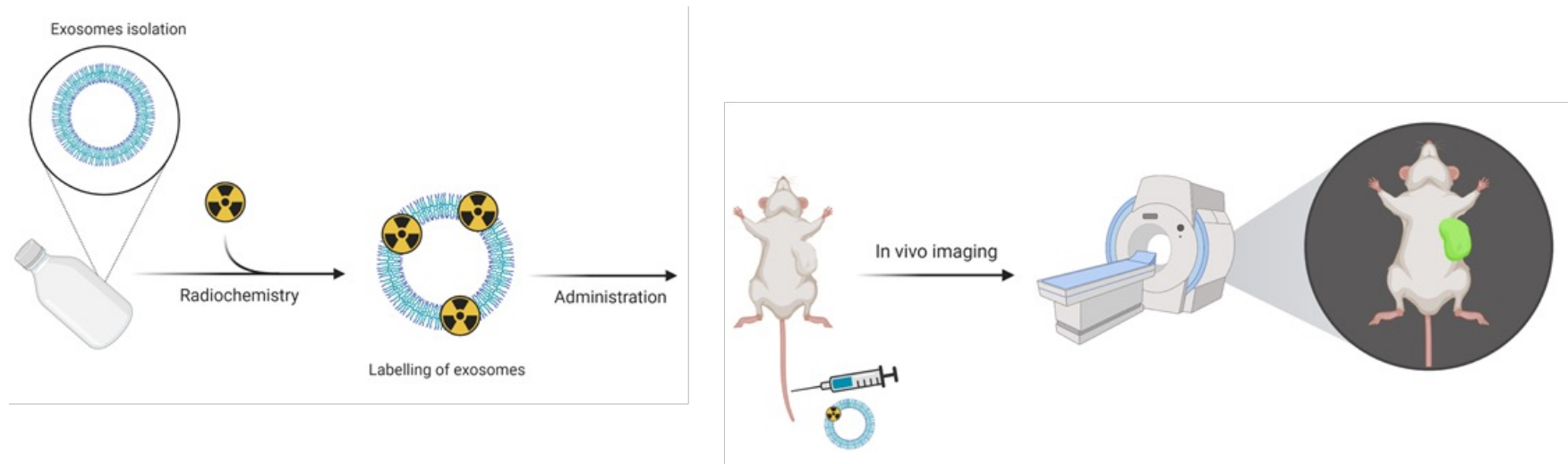


Take home message

Labelling of exosomes has allowed us to confirm not only the diagnostic capacity of the nanoparticles in tumor models but also the evaluation of the biological behavior for further use as DDS or imaging agent

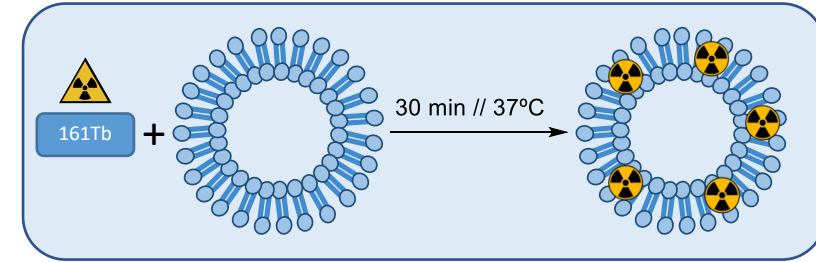
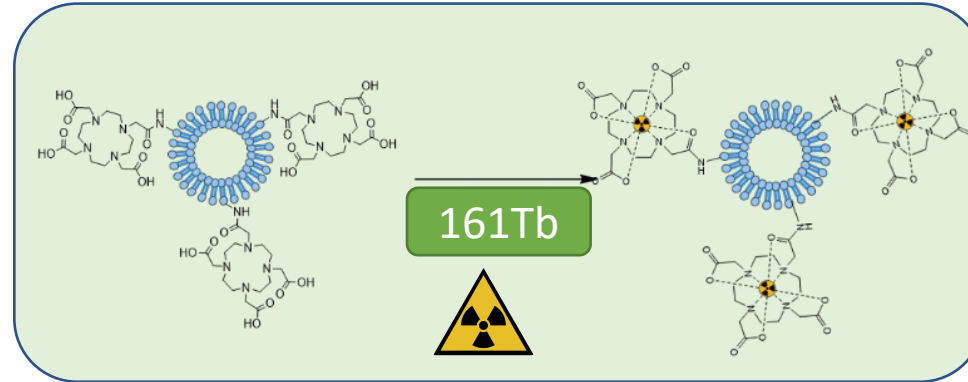
PRISMAP project

Development of new radiotheragnostic agents based on natural nanoparticles (exosomes) radioactively labeled with the novel therapeutic and diagnostic isotope Terbium 161 (^{161}Tb).

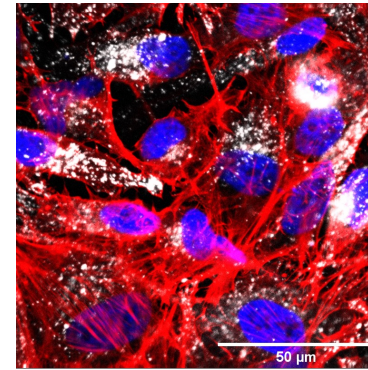
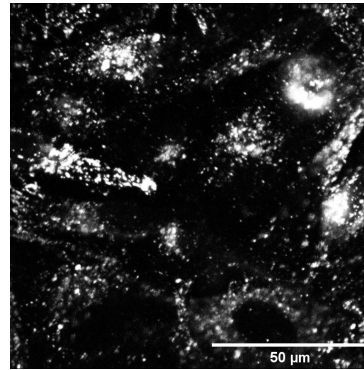


PRISMAP project

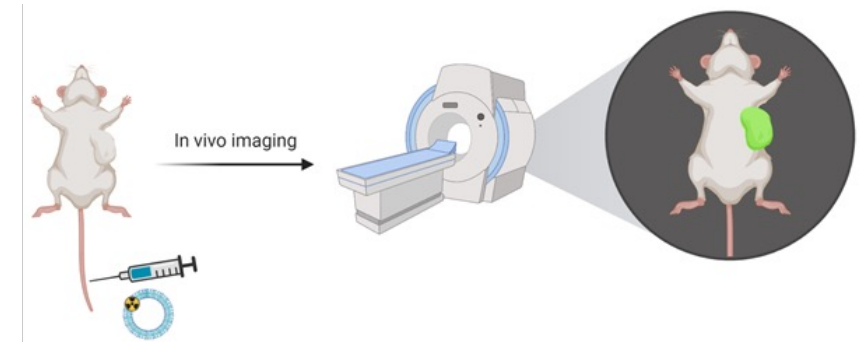
1.- Labelling ^{161}Tb



2.- *In vitro* (^{161}Tb)



3.- *In vivo* (^{161}Tb)



PRISMAP project

How did Prisma come into our life?

Previous studies in collaboration with TUM on ^{177}Lu labelling of exosomes → Dr. Weber and Dr. Kossatz

What does the Prisma consortium offer us?

- Limited access to cyclotrons and new radionuclides
- New field of research in Spain
- PRISMAP: ideal platform for the development of new lines of research

Acknowledgments



Molecular probes Lab

María Isabel González

Dra. Ana Santos

Mario González

Desiré Herreros

Gorka Sobrino

Elena Aguilera

Virginia Albaladejo

Rana Alaawar

Sara Jorqueras



Imaging Core IiSGM

Prof. Manuel Desco

Yolanda Palomares

Alexandra de Francisco

Maria Felipe

Lorena Cussó

TUM (NM Dept)

Dr. Wolfgang Weber

Dr. Susanne Kossatz



This work has been supported by the Comunidad de Madrid, projects “Y2018/NMT-4949 (NanoLiver-CM)” and “S2017/BMD-3867 (RENIM-CM)”, co-funded by European Structural and Investment Fund. This study has been also funded by Instituto de Salud Carlos III through the project “PI20/01632”, co-funded by European Regional Development Fund (ERDF), “A way to make Europe”. The CNIC is supported by Instituto de Salud Carlos III (ISCIII), Ministerio de Ciencia e Innovación (MCIN) and the Pro CNIC Foundation. A. Santos-Coquillat funded by Instituto de Salud Carlos III, co-funded by European Social Fund “Investing in your future” (Sara Borrell Fellowship grant CD19/00136). Scheme of reaction and parts of the figures were created with BioRender.com.

Fondo Europeo de Desarrollo Regional (FEDER)

“A way to make Europe”

Fondo Social Europeo (FSE)

“Investing in your future”



Selective oncological theragnostic based on radioactively labeled exosomes

Beatriz Salinas Rodríguez

Molecular probes Lab
Biomedical imaging and instrumentation Group
Instituto de investigaciones Sanitarias Gregorio Marañón.

Biomedical engineering dept, Universidad Carlos III Madrid.

Advanced Imaging Unit, National Center of Cardiovascular Diseases